

Donahaye, E.J., Navarro, S. and Leesch J.G. [Eds.] (2001) Proc. Int. Conf. Controlled Atmosphere and Fumigation in Stored Products, Fresno, CA. 29 Oct. - 3 Nov. 2000 Executive Printing Services, Clovis, CA, U.S.A. pp. 719-725

CARBON DIOXIDE UNDER HIGH PRESSURE FOR STORED-PRODUCT PROTECTION IN TEMPERATE CLIMATES

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ABSTRACT

The exposure to carbon dioxide (CO₂) under high pressure is a new control method in the food processing industry. Agricultural raw products, such as cereals, nuts or dried fruits require rapid disinfection prior to storage. The organic food branch in particular needs a preventive method that does not leave chemical residues or lead to any reduction in quality. High-pressure industrial plants work with CO₂ under pressure of 10 bar - 40 bar for a few hours. Under practical conditions, the problem of incomplete control of insect pests, occurs mostly at low temperature regimes and pressures below 15 bar.

Our experiments were conducted with the following caged stored-product insects at some of their developmental stages in different products and packages in 10 m³ chambers: *Plodia interpunctella*, *Stegobium paniceum*, *Tribolium confusum*, *Sitophilus granarius*, *Ephestia kuehniella*, *Cryptolestes ferrugineus*, *Cryptolestes turcicus*, *Trogoderma granarium*, as well as the parasitic wasp *Lariophagus distinguendus*. The results showed that at low temperatures of about 10°C at 10 bar to 15 bar of CO₂ the exposure period required for complete control varied with the product surface and the packaging material. At 15 bar and 8 to 10°C, 100% mortality was not achieved within ten hours for some of the species tested.

The rapidity at which the gas reached even distribution was found to depend on the type and mass of the product. The experiments showed that to ensure complete control it is necessary to identify the pest and classify its susceptibility prior to CO₂/high pressure treatment. The practical exposure time must also take into account the type and temperature of the product.

INTRODUCTION

Stahl and Rau (1985) and Stahl *et al.*, (1985) described a new process for residue free insect pest control by using CO₂ under high pressure. Mitsura *et al.*, (1973) were the first to report on the effects of this treatment against stored product mites. It has been found that the quality of the treated products is not disadvantageously influenced when the depressurization time is appropriately adjusted (Gerard *et al.*, 1988; Pohlen *et al.*, 1989). The growing public pressure

against the presence of insecticidal residues in food and the impending ban on the use of methyl bromide has led to investigations on alternatives for pest control. The acceptance of this new high pressure/CO₂ approach is supported by the extremely short lethal exposure time in the range of minutes or a few hours (Prozell and Reichmuth 1990 and 1991; Nakakita and Kawashima 1994; Reichmuth and Wohlgemuth 1994; Prozell *et al.* 1997). In Germany the organic food branch in particular, uses this rapid method for disinfestations of their products as a standard procedure. The objective of these experiments was to examine the efficacy of this technology during exposures at a relatively low temperature of 10°C.

MATERIAL AND METHODS

High pressure facility

All experiments were conducted in high-pressure installations of the CARVEX company (Fig. 1). The volume of the pressure chamber was 9 m³. Carbon dioxide in a pressurized tank connected to the exposure chamber, provided the gas supply. Before initiating treatment, the CO₂ was warmed and then introduced into the pressure-chamber. Different pressure regimes were made available by adjustment of the regulator (Table 1).

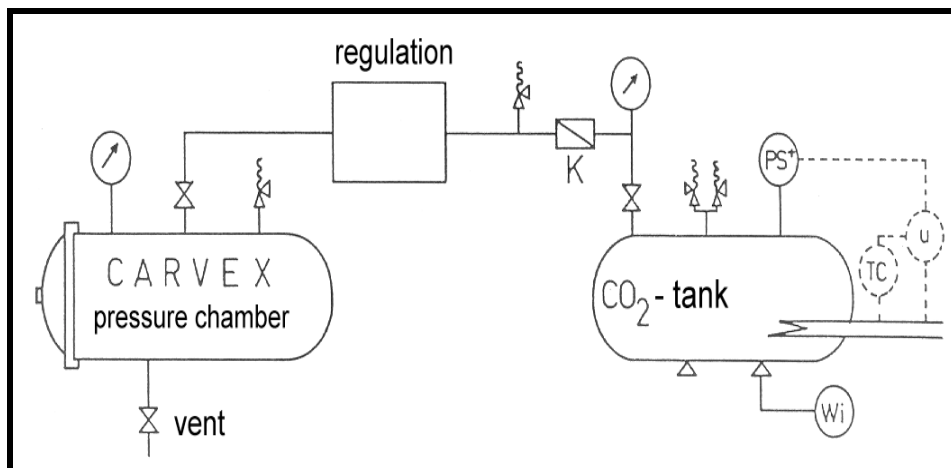


Fig. 1: Pressure chamber to hold carbon dioxide, carbon dioxide supply tank and regulation unit (Wi: balance, PS+, u, TC: supply gear to adjust temperature and pressure in the tank, K: valve) (after Gerard *et al.*, 1990).

At the end of the required exposure-period the pressure was released. The exposure time included the time to build up the targeted pressure. The time for depressurization ranged from 14 to 25 min.

TABLE 1
Pest species, carbon dioxide pressure, temperature and exposure period of the experiments

Insect species	Pressure in bar	Temp °C	Exposure time in h
<i>Sitophilus granarius</i>	15	0 to 2	11.5
	20	4	8
	15	5	15
<i>Lariophagus distinguendus</i>	15	0 to 2	11.5
	20	4	8
	15	5	15
<i>Tribolium confusum</i>	15	0 to 2	11.5
	20	4	8
	15	/5	15
<i>Ephestia kuehniella</i>	15	0 to 2	11.5
	20	4	8
	15	5	15
<i>Cryptolestes ferrugineus</i>	15	0 to 2	11.5
	20	4	8
	15	5	15
<i>Cryptolestes turcicus</i>	15	0 to 2	11.5
	15	0 to 5	15
	20	/0 to 4	8
<i>Stegobium paniceum</i>	15	0 to 2	11.5
	15	0 to 5	15
	20	0 to 4	8
<i>Trogoderma granarium</i>	15	0 to 2	11.5
	15	0 to 5	15
	20	0 to 4	8
<i>Plodia interpunctella</i>	15	0 to 2	11.5
	15	0 to 5	15
	20	0 to 4	8

Insects

Experiments were performed using all developmental stages of a mixture of several pest species (Table 1). The insects were introduced inside stainless steel wire mesh cages (10 cm length, 1 cm diameter) fitted with rubber stoppers. Prior to treatment, separate cages were distributed to different positions inside the chamber. Two exposure profiles were examined: in one, the cages were placed at the centre of a metal bucket containing flour; in the other, the cages were placed at the centre of a 'big-bag' of 1 m³ capacity, made from webbed PP mesh and containing herbal tea. The chamber was then closed and pressurized with CO₂. At the end of the treatment the pressure was released, the cages removed, and the samples were transferred to an incubator at 26°C and 75% r.h., and observed weekly for detection of survivors during the following 14 weeks. Control samples of insects were prepared similarly but not subjected to treatment.

RESULTS

High-pressure treatment in big-bags

High pressure treatments in 'big-bags' caused 100% mortality of *Plodia interpunctella*, *Stegobium paniceum* and *Lariophagus distinguendus*. (See Table 2 for comprehensive details of results).

High-pressure treatment in flour

More survivors were found in test cages placed in the flour. Only *Sitophilus granarius*, *Tribolium confusum* and *Stegobium paniceum* were completely controlled in all experiments. (See Table 2 for all results)

Control insects

All insects in the untreated control samples developed normally.

DISCUSSION AND CONCLUSIONS

The results of insect mortality presented in these experiments are similar to those with *S. granarius* described by Prozell and Reichmuth (1990 and 1991). The toxic action of inert gases under increased pressure was first described by Ferguson and Hawkins (1949), and later by Johnson and Quastel (1953), and Carpenter (1954). They mentioned narcotic effects after treatment with these gases. Insect death presumably occurs during treatment under high pressure as a consequence of prolonged and intense narcosis. Destruction of cell membranes during decompression also causes severe damage (Ulrichs 1994). Prozell *et al.*, (1997) stated that the speed of distribution of the CO₂ under pressure seems to depend on the type and density of the treated product.

The presented investigation into the rapidity at which CO₂ distributes itself through the product revealed similar results. Previous work showed that at first, compressed air remained in the centre of the product, surrounded by CO₂ under pressure. The initial difference in pressure is not sufficient to quickly remove all the residual air from the interstitial space within the product. On the other hand, pressurization of the air alone does not control insect pests in a short time (Prozell and Reichmuth 1991). Later during exposure, the CO₂ content increased also in the centre of the product mainly due to relatively slow diffusion. Four time phases of penetration can be discussed which follow the classical transport phenomena (Bird *et al.* 1960). The time required to obtain the necessary CO₂ content to control insect pest can be delayed inside compressed products, because a longer time for uniform distribution will be required (Prozell *et al.* 1997). The results presented here show that it is advantageous and even necessary to identify and classify the sensitivity of the pest to treatment, the nature of the product to be treated, the exposure temperature and possibly the existence of developmental stages prior to undertaking a high-pressure treatment. Complete mortality can be achieved more slowly in 'big-bags' than in small containers with flour. Mortality rates depend also on the size of the insects and their developmental stages and whether infestation occurs inside or outside the particles of a treated commodity (Ulrichs *et al.*, 1997a and 1997b). With all this information at hand, the required exposure time can be adjusted accordingly. In contrast to the conventional insecticides and toxic fumigants this treatment can be used as a preventive method to ensure pest free food and feed, without leaving chemical residues.

TABLE 2

Results of carbon dioxide/high pressure treatment of various developing stages of various stored product pest insects in flour and big bags, (x = survivors, 0 = no survivors)

Insect species	Product/ container	Exposure time in h	Temp °C	Survivors
<i>Sitophilus granarius</i>	flour	11.5	0 to 2	0
	flour	11.5	0 to 2	0
	flour	15	5	0
	big bag	15	5	0
	big bag	8	4	0
	flour	6	12 to 15	0
	big bag	6	12 to 15	0
	flour	7	12	0
	big bag	7	12	X
<i>Lariophagus distinguendus</i>	flour	11.5	0 to 2	X
	big bag	11.5	0 to 2	0
	flour	8	4	X
	big bag	8	4	0
	flour	15	5	X
	big bag	15	5	X
<i>Tribolium confusum</i>	flour	11.5	0 to 2	X
	flour	8	4	0
	flour	15	5	0
<i>Ephestia kuehniella</i>	Flour	11.5	0 to 2	X
	flour	8	4	0
	flour	15	5	0
<i>Cryptolestes ferrugineus</i>	big bag	11.5	0 to 2	0
	big bag	8	4	0
	flour	8	4	X
	flour	15	5	X
	big bag	15	5	X
<i>Cryptolestes turcicus</i>	big bag	6	12 to 15	X
	flour	6	12 to 15	X
	big bag	7	12	X
	flour	7	12	X
	big bag	10	8 to 12	X
	flour	10	8 to 12	X
<i>Stegobium paniceum</i>	big bag	6	12 to 15	0
	flour	6	12 to 15	0
	big bag	7	12	0
	flour	7	12	0
	big bag	10	8 to 12	0
	flour	10	8 to 12	0
<i>Trogoderma granarium</i>	big bag	6	12 to 15	X
	flour	6	12 to 15	X
	big bag	7	12	X
	flour	7	12	X
	big bag	10	8 to 12	0
	flour	10	8 to 12	X
<i>Plodia interpunctella</i>	big bag	6	12 to 15	0
	flour	6	12 to 15	X
	big bag	7	12	0
	flour	7	12	X
	big bag	10	8 to 12	0
	flour	10	8 to 12	X

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