

Donahaye, E.J., Navarro, S. and Leesch J.G. [Eds.] (2001) *Proc. Int. Conf. Controlled Atmosphere and Fumigation in Stored Products*, Fresno, CA. 29 Oct. - 3 Nov. 2000, Executive Printing Services, Clovis, CA, U.S.A pp. 757-762

COMPARATIVE RESPONSES OF THREE PSOCID SPECIES (PSOCOPTERA: LIPOSCOLIDIDAE) TO FIVE FUMIGANTS

J. RIUDAVETS,¹ K.A. DAMCEVSKI^{2*} AND P.C. ANNIS²

¹*Institut de Recerca i Tecnologia Agroalimentàries (IRTA). 08348 Cabrils (Barcelona)
Spain*

²*Stored Grain Research Laboratory. CSIRO Entomology. Canberra ACT 2601 Australia
[*e-mail: katherine.damcevski@ento.csiro.au]*

ABSTRACT

The responses of three different psocid species, *Liposcelis bostrychophila*, *L. decolor* and *L. taeta*, were compared when treated with the fumigants carbonyl sulfide, carbon disulfide, ethyl formate, carbon dioxide and phosphine. Eggs of psocids were exposed to one applied concentration of these gases in desiccators and mortality recorded at three different exposure periods and at three different temperatures. The three psocid species had different tolerances to each of the 5 fumigants tested and the pattern of response between species varied depending on the fumigant. Under the combinations of concentrations, times and temperatures used, the response varied widely between these psocid species. The least variability of control overall was achieved with ethyl formate. Overall, *L. taeta* was the most difficult to control.

INTRODUCTION

Complaints due to psocid infestations are currently increasing in stored commodities around the world (Turner, 1994; Rees, 1998). Large populations often develop after treatments with controlled atmosphere (CA) or fumigation. Among possible causes commonly reported are: fumigation failure, short treatment times, resistance to fumigants and selective removal of natural enemies (Pike, 1994; Santoso *et al.*, 1996; Roesli *et al.*, 1997).

A number of potential new fumigants are being currently studied for the control of stored product pests (Desmarchelier, 1998; Reichmuth, 1999). Despite the importance of psocids (Psocoptera), there is little information on their susceptibility to these fumigants. Phosphine (PH₃) has been found to be highly effective against *Liposcelis bostrychophila* (Badonnel), *L. decolor* (Pearman) and *L. taeta* (Pearman) (Nayak *et al.*, 1998). Eggs of *L. bostrychophila* were found to be more tolerant than adults to PH₃ (Ho and Winks, 1995) and to CO₂ (Leong and Ho, 1991). Weller and Beckett (in press) recently found that PH₃ at 10, 20 and 35 ppm, for 15 and 28 d,

controlled populations of *L. bostrychophila*, *L. tenebrosa*, and *L. antinomophila*, at 25 and 30°C.

The aim of our research was to investigate the response of three psocid species to a range of potential fumigants so that effective application rates can be recommended for each species.

MATERIAL AND METHODS

Species tested were *L. bostrychophila*, *L. tenebrosa* and *L. antinomophila*. The strain of *L. bostrychophila* was originally obtained from Gogeldrie, New South Wales, *L. tenebrosa* from Thevenard, South Australia and *L. antinomophila* from Ayr, Queensland, Australia.

Eggs were exposed to one applied concentration of each fumigant in glass desiccators (Table 1). The fumigant concentrations and times were selected to give mortalities between 80 and 100%. These selections were based largely on unpublished data with the exception of CO₂, which was based on data from Leong and Ho (1991). The ethyl formate (EtF) used was laboratory grade (920 g/L); carbon disulfide (CS₂) was laboratory grade (1,263 g/L); carbonyl sulfide (COS) was cylinderized gas (98%); CO₂ was mixed from cylinderized CO₂ (100%) and air; and PH₃ was generated by addition of aluminium phosphide tablets to 5% (v/v) aqueous sulphuric acid.

Each fumigated sample consisted of 15g of coarsely milled rolled oats compacted into a plastic or glass dish (7 cm diameter and 5 cm high) as used by Beckett (1998). The medium was conditioned in an incubator at 70% r.h. for 15 d before use. To obtain eggs for fumigation, 50 adults were isolated on a dish of media for 2 d. Duplicates of each species were exposed in the same desiccator to one applied concentration of each gas for a range of exposure times and temperatures of 15, 25 and 30°C (Table 1). Gas concentrations were measured during the exposures by gas chromatography, within an hour of dosing and at approximately 24-h intervals, or less for the shorter exposures. The reduction in numbers of emergent nymphs was assessed.

TABLE 1
Fumigants and dosages tested

Fumigant	Concentration	Exposure times
Ethyl formate	25 mg/L	6, 12, 24, 48 hours
Carbon disulfide	12 mg/L	1, 2, 3 days
Carbonyl sulfide	15 mg/L	1, 2, 3 days
Carbon dioxide	40% (~12%O ₂)	2, 3, 4 days
Phosphine	0.03 mg/L (20 ppm)	2, 3, 4 days

RESULTS AND DISCUSSION

Analysis and sorption of fumigants

The concentration of COS, CO₂ and PH₃ remained constant during all the exposure times and at all temperatures tested, indicating that for these three fumigants no significant sorption or breakdown occurred. However, the concentrations of CS₂ and EtF declined during the experiment (Figs. 1, 2). This was probably due to sorption by the medium and in the case of EtF also due to breakdown of the chemical. Decline in concentration of EtF and CS₂ was greater at 15°C than at 25 and 30°C.

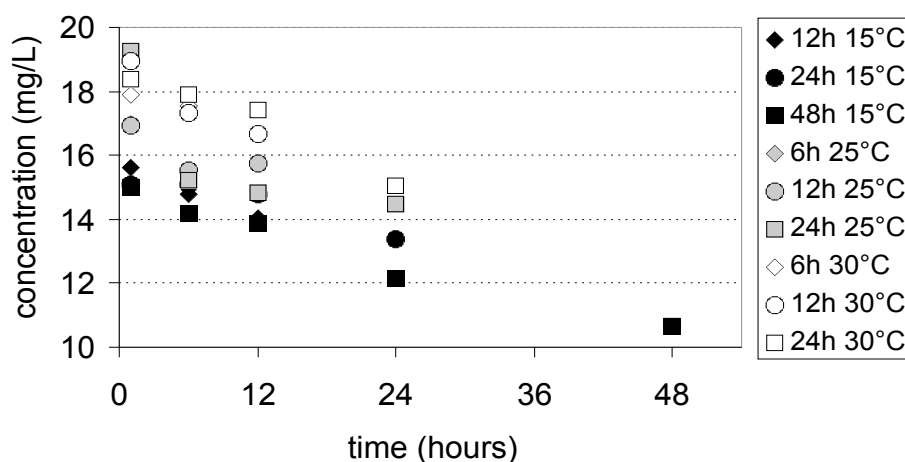


Fig. 1. Changes in ethyl formate concentration during fumigation. Ethyl formate applied at 25 mg/L.

Insect mortality

L. bostrychophila, *L. paeta* and *L. decolor* showed different tolerances to each of the 5 fumigants tested and the pattern of response varied depending on the fumigant, (Tables 2, 3). For EtF, all three species responded similarly to the combinations of concentrations, times and temperatures tested. However, *L. decolor* was the most tolerant to this fumigant at 30°C, and *L. bostrychophila* was less tolerant than the other two species at 25°C. For CS₂, *L. bostrychophila* was more tolerant than *L. decolor* and this latter species more tolerant than *L. paeta* at all three temperatures tested.

By contrast, *L. paeta* was the most tolerant species to all dosages and temperatures tested of the other three fumigants. *L. decolor* was the least tolerant of all three species to COS, CO₂ and PH₃. The pattern of response to these fumigants did not vary over the temperature range tested.

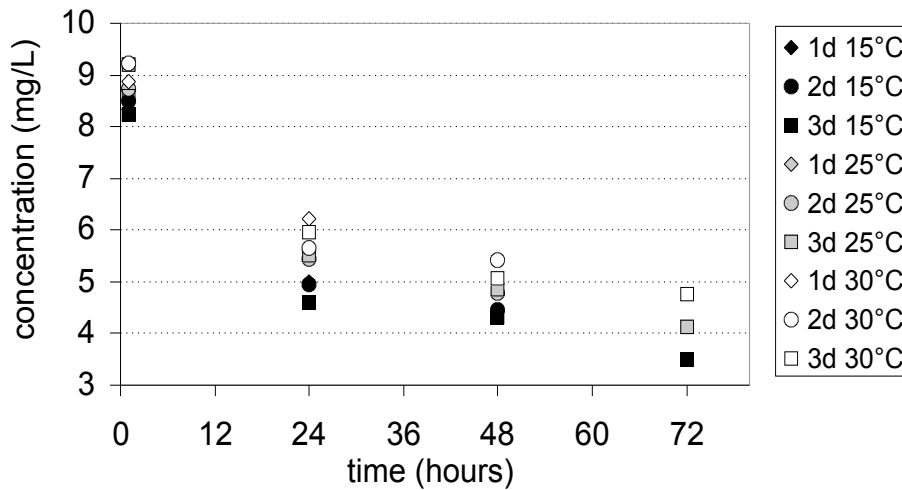


Fig. 2. Changes in carbon disulfide concentration during fumigation. Carbon disulfide applied at 12 mg/L.

TABLE 2
Percentage of progeny reduction of *L. bostrychophila* (BOS), *L. decolor* (DEC)
and *L. taeta* (PAE) when treated with ethyl formate and carbon disulfide

Fumigant	Temperature (°C)	Exposure time (hours)	% Progeny reduction		
			BOS	DEC	PAE
Ethyl formate 25 mg/L	15	12	90 - <100	90 - <100	90 - <100
		24	100	100	100
		48	100	100	100
	25	6	90 - <100	90 - <100	90 - <100
		12	100	90 - <100	90 - <100
		24	100	100%	100%
	30	6	100	90 - <100	100
		12	100	100	100
		24	100	100	100
Carbon disulfide 12 mg/L	15	24	75 - <90	90 - <100	90 - <100
		48	90 - <100	90 - <100	90 - <100
		72	90 - <100	100	100
	25	24	75 - <90	90 - <100	100
		48	90 - <100	100	100
		72	90 - <100	100	100
	30	24	90 - <100	90 - <100	100
		48	90 - <100	100	100
		72	100	100	100

TABLE 3
Percentage of progeny reduction of *L.Bostrychophila* (BOS), *L.Decolor* (DEC) and *L.Paeta* (PAE) when treated with carbonyl sulfide, carbon dioxide and phosphine

Fumigant	Temperature °C	Exposure time (hours)	% Progeny reduction		
			BOS	DEC	PAE
Carbonyl sulfide 15 mg/L	15	24	75 - <90	75 - <90	<75
		48	90 - <100	90 - <100	<75
		72	90 - <100	100	75 - <90
	25	24	75 - <90	90 - <100	<75
		48	90 - <100	100	90 - <100
		72	100	100	90 - <100
	30	24	90 - <100	90 - <100	75 - <90
		48	90 - <100	100	90 - <100
		72	100	100	90 - <100
Carbon dioxide 40% (12%O ₂)	15	48	75 - <90	90 - <100	<75
		72	75 - <90	90 - <100	75 - <90
		96	90 - <100	100	90 - <100
	25	48	90 - <100		75 - <90
		72	90 - <100	100	90 - <100
		96	90 - <100	100	90 - <100
	30	48	90 - <100	90 - <100	75 - <90
		72	90 - <100	100	90 - <100
		96	90 - <100	100	90 - <100
Phosphine 0.03 mg/L (20 ppm)	15	48	75 - <90	75 - <90	<75
		72	75 - <90	90 - <100	<75
		96	90 - <100	90 - <100	<75
	25	48	90 - <100	90 - <100	<75
		72	90 - <100	90 - <100	<75
		96	90 - <100	100	<75
	30	48	90 - <100	100	<75
		72	90 - <100	100	75 - <90
		96	90 - <100	100	75 - <90

These results indicate that within the species and strains tested, *L.Paeta* was the most tolerant species to PH₃, COS and CO₂, *L.Bostrychophila* was the most tolerant species to CS₂ and *L.Decolor* was the most tolerant to EtF for the combinations of concentrations, times and temperatures tested. However, there can be just as much variation in response between strains of a given species as between individual species as observed by Weller and Beckett (in press). Therefore, in order to control psocids it may be necessary to establish which species, and possibly which strains, are present in an infestation and apply the correct fumigant at the correct rate, although it would probably be sufficient to ensure that recommended applications cover the most tolerant species.

ACKNOWLEDGEMENTS

This research was supported by a grant from the Comissionat per a Universitats i Recerca del Departament de la Presidència de la Generalitat de Catalunya and by S.E. de Carburos Metálicos S.A., and by participants to the Stored Grain Research Laboratory Agreement.

REFERENCES

- Beckett, S. (1998) Treating psocids with heat: an alternative grain disinfestation treatment for a new pest. In: *Stored Grain in Australia. Proc. Australian Postharvest Technical Conference*, (Edited by Banks, H.J, Wright, E.J and Damcevski, K.A.), Canberra, 26-29 May 1998, 334-337.
- Desmarchelier, J.M. (1998) Potential new fumigants. In: *Stored Grain in Australia. Proc. Australian Postharvest Technical Conference*, (Edited by Banks, H.J, Wright, E.J and Damcevski, K.A.), Canberra, 26-29 May 1998, 133-137.
- Ho, S.H. and Winks, R.G. (1995) The response of *Liposcelis bostrychophila* Badonnel and *L. entomophila* (Enderlein) (Psocoptera) to phosphine. *J. stored Prod. Res.*, **31**, 191-197.
- Leong, E.C.W. and Ho, S.H. (1991) Research on *Liposcelis bostrychophila* Badonnel and *Liposcelis entomophilus* (Enderlein) (Psocoptera: Liposcelididae). In: *Proc. 14th ASEAN Seminar on Grain Postharvest Technology*, (Edited by Naewbanij, J.O., and Manilay, A.A.), Manila, Philippines, 317-327.
- Nayak, M.K., Collins, P.J. and Reid, S.R. (1998) Efficacy of grain protectants and phosphine against *Liposcelis bostrychophila*, *L. entomophila* and *L. paeta* (Psocoptera: Liposcelididae). *J. Econ. Entomol.*, **91**, 1208-1212.
- Pike, V. (1994) Laboratory assessment of the efficacy of phosphine and methyl bromide fumigation against all life stages of *Liposcelis entomophilus* (Enderlein). *Crop Protection*, **13**, 141-145.
- Rees, D. (1998) Psocids as pests of Australian grain storages. In: *Stored Grain in Australia. Proc. Australian Postharvest Technical Conference*, (Edited by Banks, H.J, Wright, E.J and Damcevski, K.A.), Canberra 26-29 May 1998, 46-51.
- Reichmuth, C. (1999) Fumigation for pest control in stored product protection. In: *Proc. 7th Int. Working Conf. on Stored-product Protection*, (Edited by: Zuxun, J., Quan, L., Yongsheng, L., Xianchang, T. and Lianghua, G.), Beijing. Sichuan Publishing House of Science & Technology, Chengdu, China, **1**, 311-318.
- Roesli, R., Jones, R. and Rees, D.P. (1997) Behind the explosion of *Liposcelis* populations. In: *Proc. Symposium on Pest Management for Stored Food and Feed*. 5-7 September 1995, Bogor, Indonesia. SEAMEO BIOTROP, Special Publication 59, 127-129.
- Santoso, T., Sunjaya, Dharmaputra, O.S., Halid, H. and Hodges, R.J. (1996) Pest management of psocids in milled rice stores in the humid tropics. *Int. J. Pest Management*, **42**, 189-197.
- Turner, B.D. (1994) *Liposcelis bostrychophila* (Psocoptera: Liposcelididae), a stored pest in the UK. *Int. J. Pest Management*, **40**, 179-190.
- Weller, G. and Beckett, S. (2001). Can SIROFLO[®] label rates control psocid infestations? In: *Proc. Australian Postharvest Technical Conference*, Adelaide, 1-4 Aug. 2000.